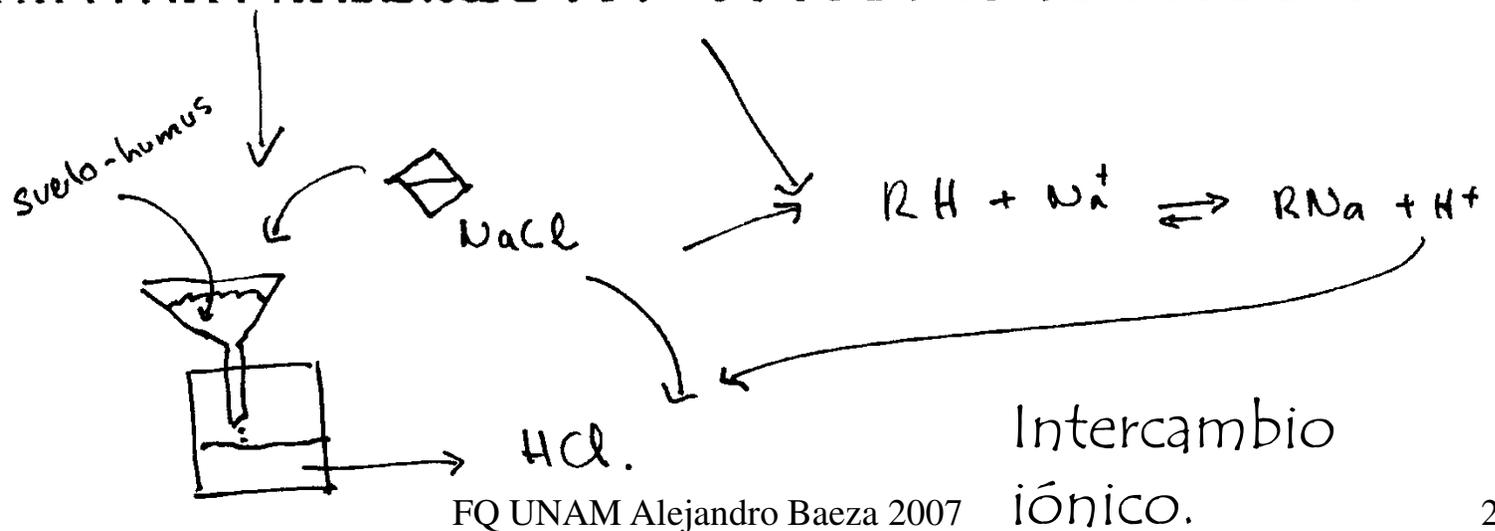
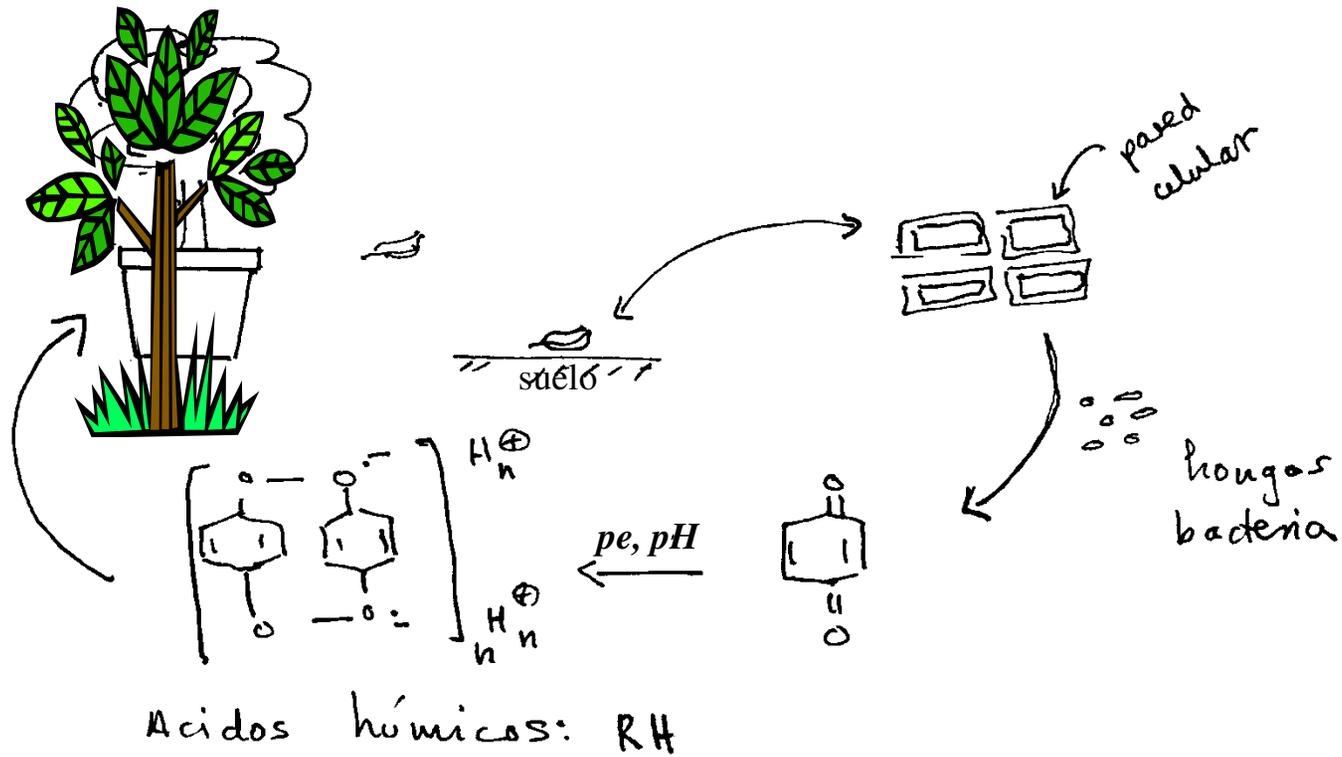


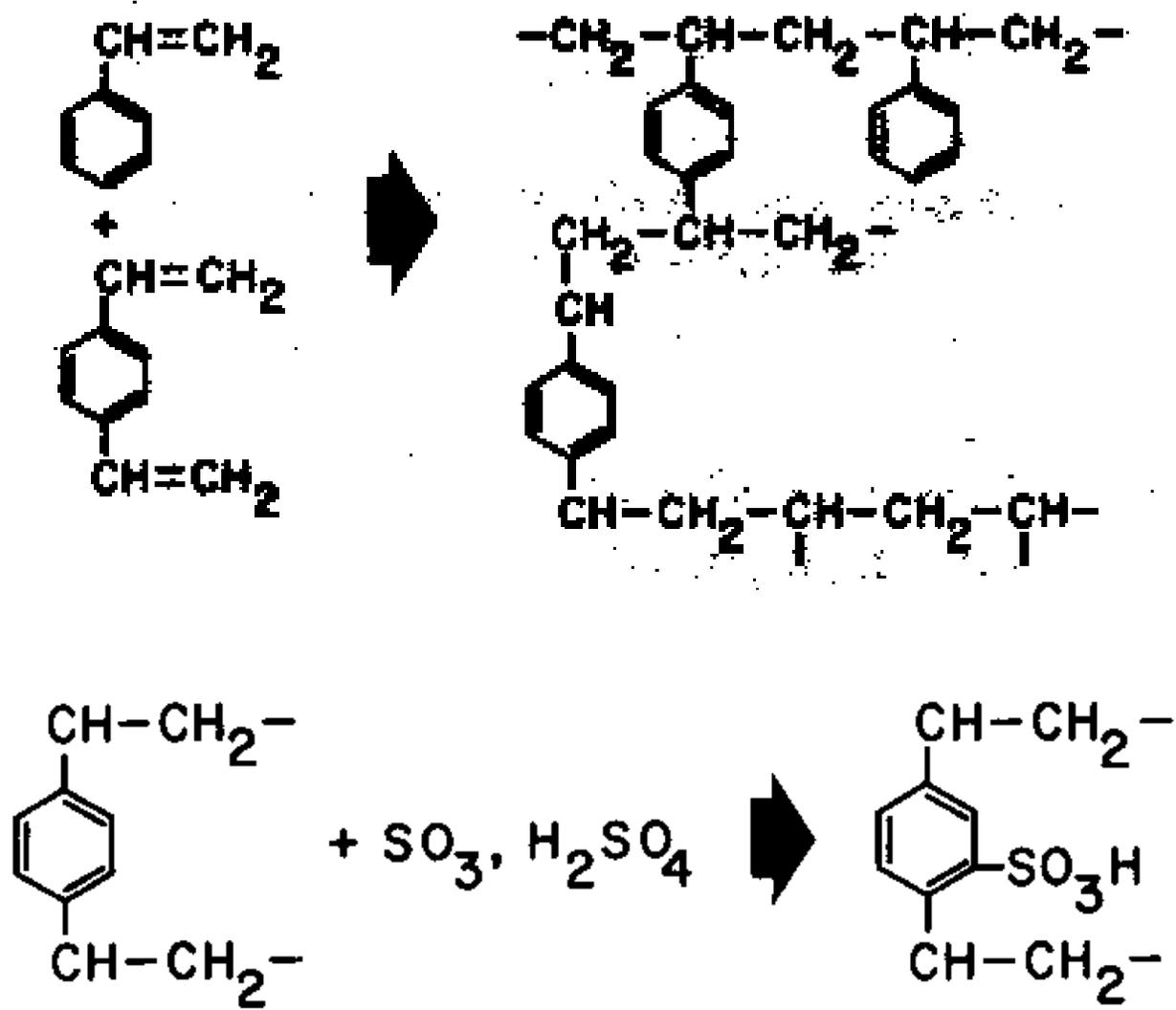
Química Analítica I (1402)

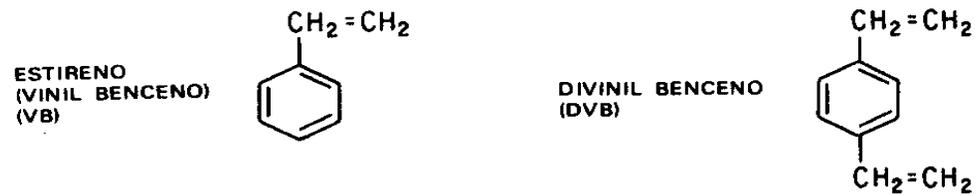


Resinas intercambiadoras de iones

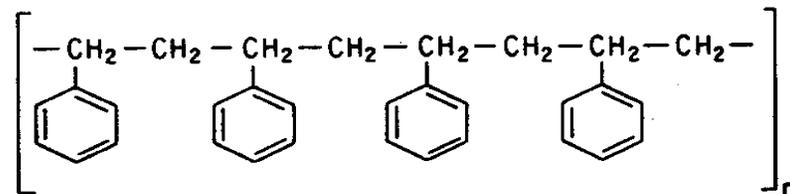
(Aspectos generales)







(2) POLIMERIZACION DEL ESTIRENO EN POLIESTIRENO



(3) COPOLIMERIZACION DEL ESTIRENO Y DIVINIL BENCENO. MOSTRANDO LA FUNCION QUE DESEMPEÑA EL DVB EN EL ENTRECruzAMIENTO DE LAS CADENAS DEL POLIESTIRENO.

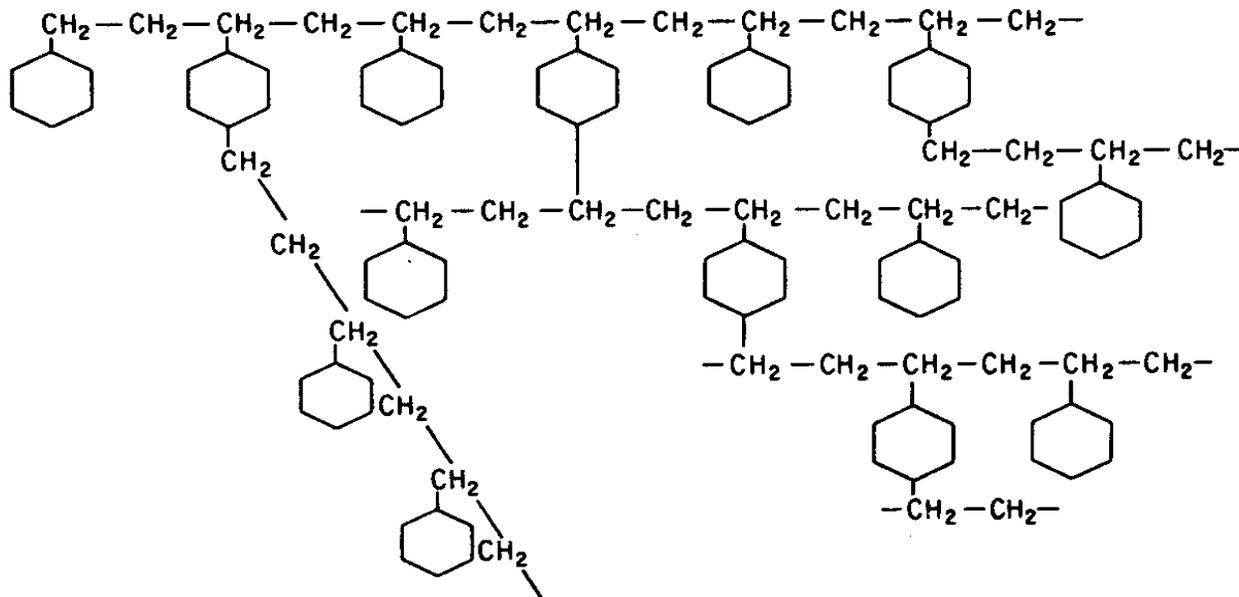


Figura 12.3 Producción de resinas intercambiadoras de iones estables mediante copolimerización.

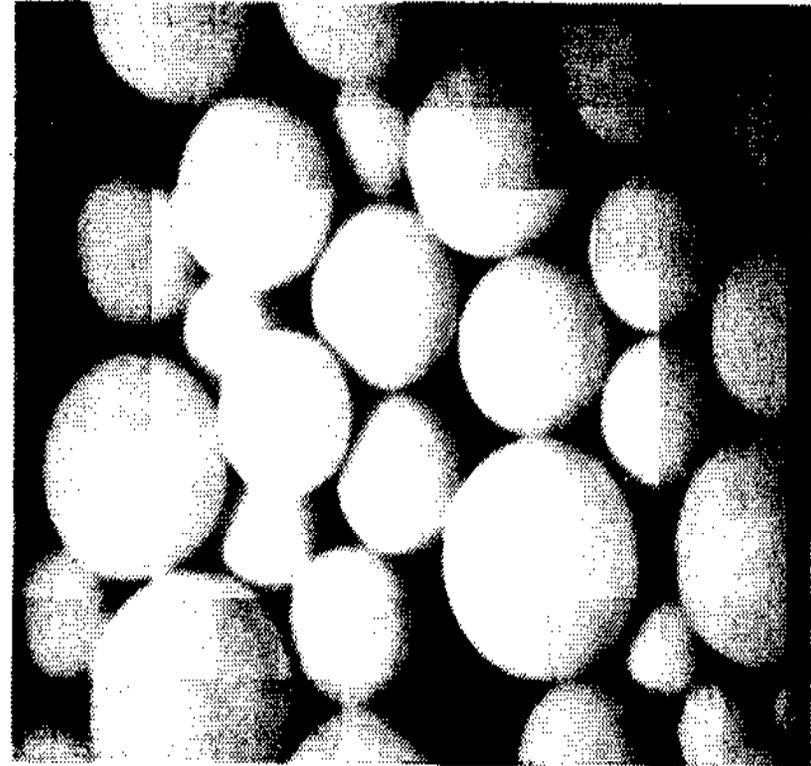
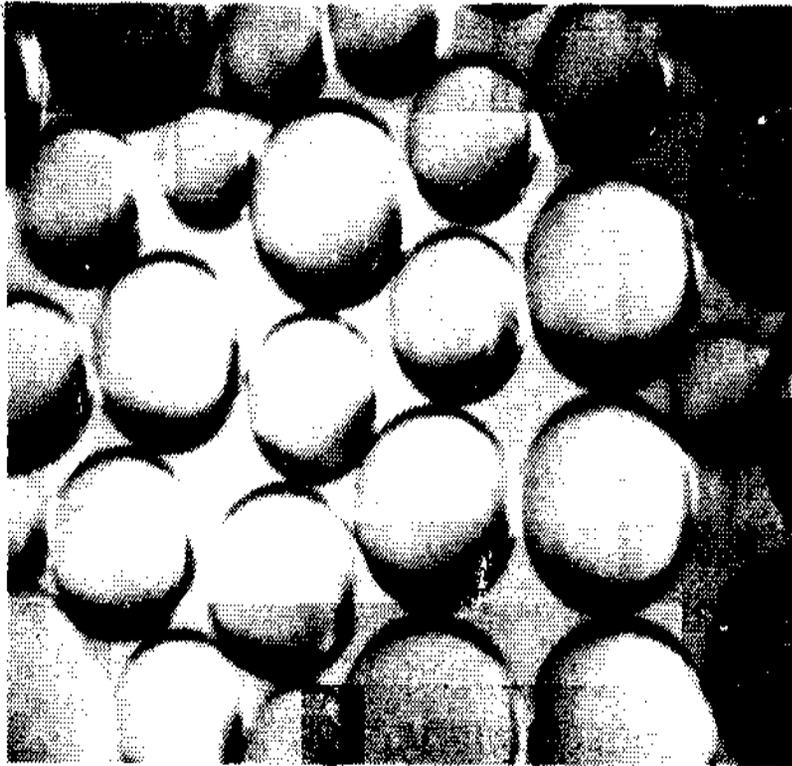


Figura 12.2 Los intercambiadores comerciales típicos pertenecen a dos estructuras generales, del tipo de gel (izquierda) y del tipo de macroporo (derecha). Estas son ampliificaciones.

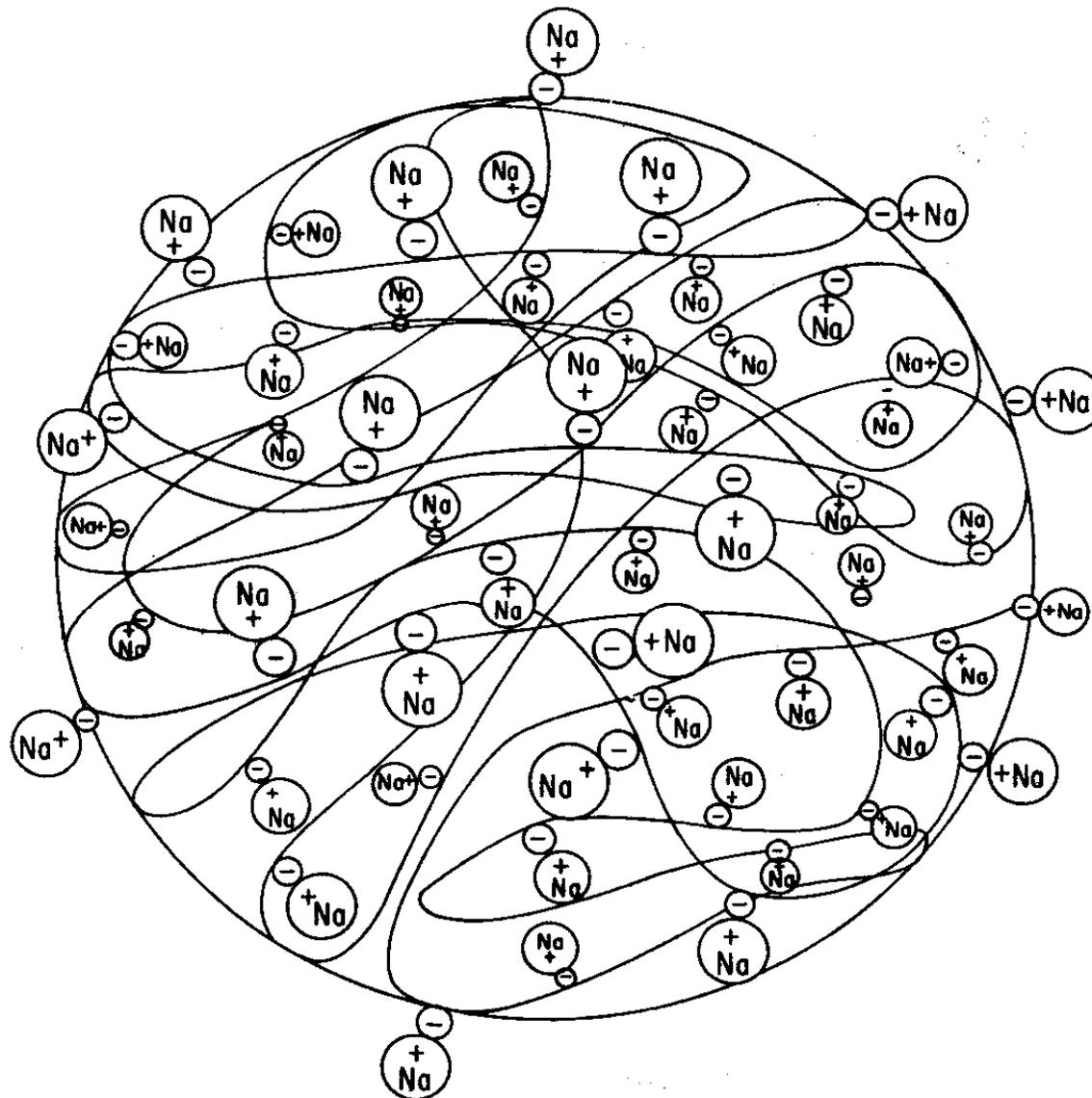
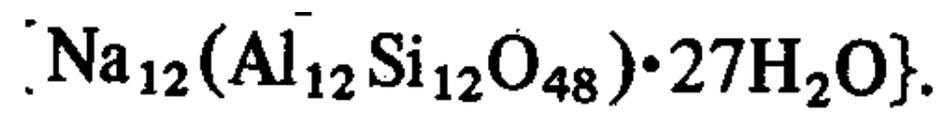
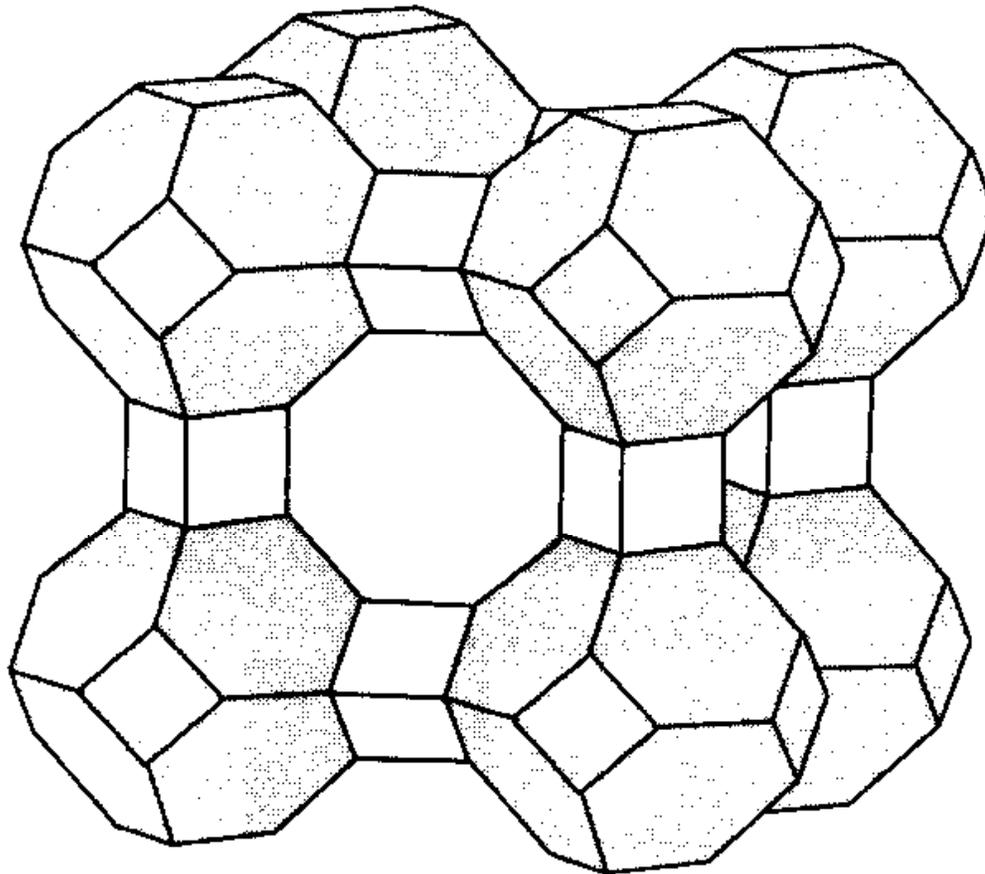
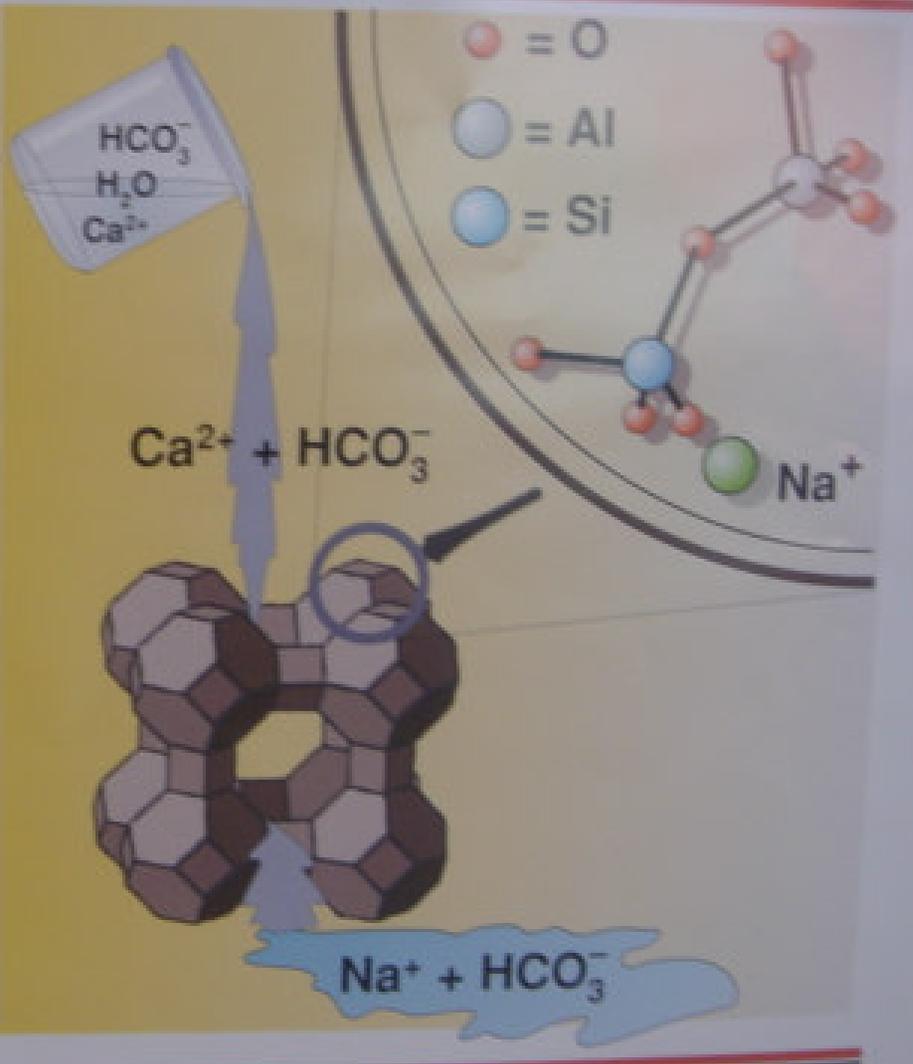
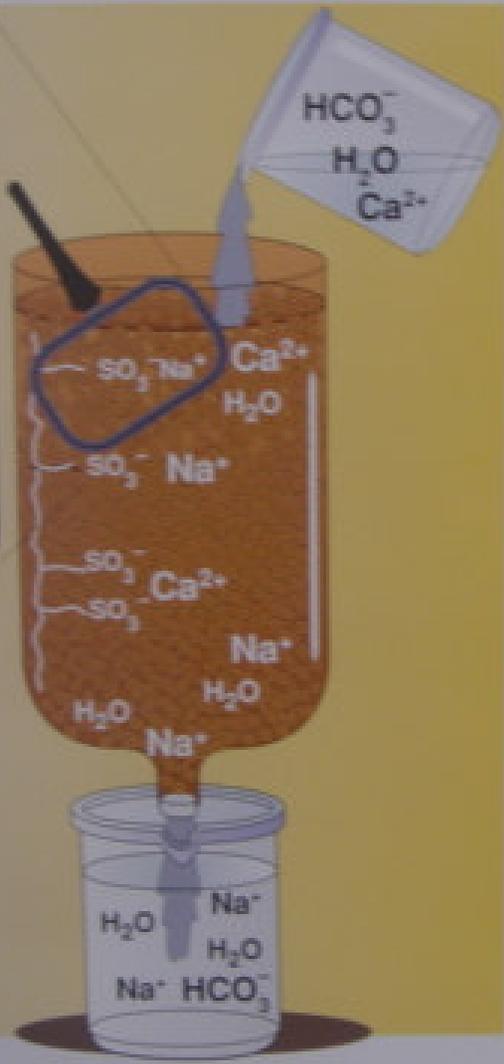
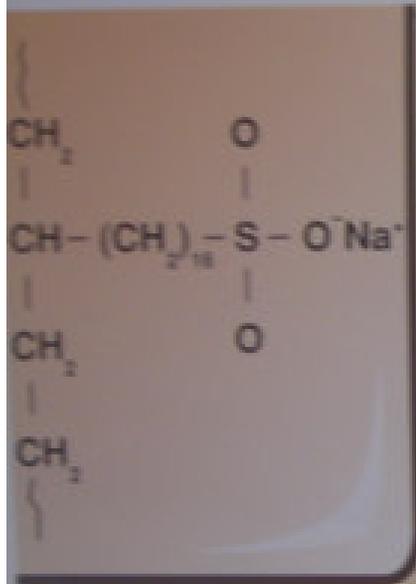


Figura 12.1 Modelo de un intercambiador de cationes, mostrando los sitios de intercambio negativamente cargados sobre el esqueleto que sostiene los iones de sodio como uvas en un racimo.

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L 14

* Aplicaciones analíticas del intercambio iónico.

valoraciones titulométricas bifásicas
(*desarrollo analítico farmacéutico*)

separaciones analíticas
(*aminoácidos, cationes*)

Aplicaciones sintéticas del intercambio iónico



separaciones preparativas
(*productos naturales, Plutonio*)

desmineralización de agua

síntesis inorgánica

Vehículo de medicamentos

TABLE XIX
Anion Exchange Determination of Alkaloidal Salts

| Compounds analyzed | Sample size | Resin, (OH ⁻) form | Charge | Sample solvent | Eluent | Titrant | Indicator |
|-----------------------------|--------------|--------------------------------|-------------------------|--------------------------|---------------------------------------|-----------|---------------------------|
| Salts of Czech. Pharm. 2 | 100 mg. | Amberlite IR-4B | 1 × 10 cm. | Hot 80% ethanol (10 ml.) | Hot 80% ethanol (50 ml.) | 0.1N HCl | Methyl red-methylene blue |
| Various alkaloidal salts | 100-200 mg. | Amberlite IR-4B | 0.5 × 5 cm. | 95% Ethanol (20 ml.) | Hot 95% ethanol (50 ml.) ^b | 0.1N HCl | Methyl red-methylene blue |
| | 10-30 mg. | Amberlite IR-4B | 0.8 × 3.5 cm. (2.5 gm.) | Hot 90% ethanol (10 ml.) | Hot 90% ethanol (20 ml.) | 0.01N HCl | Potentiometrically |
| | 0.5-1.0 meq. | Amberlite IRA-400 | 0.8 × 7.5 cm. | 50% Ethanol | 50% Ethanol (50 ml.) | 0.1N HCl | Bromophenol blue |
| Antihistamine salts | 0.25 meq. | Dowex 2 | 1 × 13.5 cm. | 70% Ethanol | 70% Ethanol | 0.1N HCl | Bromophenol blue |
| | | Amberlite IRA-410 ^c | 1 × 10 cm. | 60% Ethanol (15 ml.) | 60% Ethanol (50 ml.) | 0.1N HCl | Bromocresol green |
| Local anesthetic salts | 20-50 mg. | Amberlite IRA-400 | 8-10 gm. | 75% Ethanol (20 ml.) | Hot 95% ethanol (30 ml.) | 0.1N HCl | Methyl red |
| Narcotic salts | 75 mg. | Amberlite IR-4B | | 50% Ethanol | 75% Ethanol | 0.1N HCl | Methyl red |
| Sympathomimetic amine salts | 25 mg. | Amberlite IR-45 ^d | | Water | 75% ethanol (200 ml.) ^e | 0.1N HCl | Methyl red |

^a Weaker bases are titrated potentiometrically.

^b Methanol is used for morphine and quinine salts, and 75% ethanol for strychnine nitrate.

^c A 1 × 20 cm. charge of Amberlite IR-4B may be used.

^d Nonphenolic compounds are also determined with this method.

^e A large volume of eluent is required to remove the sample.

TABLE XX
Cation Exchange Determination of Alkaloidal Bases

| Sample | Resin | Solvent | Wash | Eluent | Determination | Refs. |
|------------------|---|-------------------------------------|--|--------------------------------------|---------------------------------------|------------------|
| Total alkaloids* | | | | | | |
| A. Belladonna | IMAC-C22 | 70% Alcohol acidulated with HCl | (1) 70% Alcohol, (2) 50% alcohol, (3) water, (4) basic NH ₄ OH-NH ₄ Cl buffer, (5) water | 1N NH ₄ OH | A. Titration, 0.1N HCl ^{b,c} | (394) |
| B. Cinchona | (H ⁺) | | | | B. Titration, 0.1N HCl ^b | |
| C. Hydrastis | | | | | C. Gravimetric | |
| D. Hyoseyamus | | | | | D. Titration, 0.1N HCl ^{b,c} | |
| E. Ipecac | | | | | E. Titration, 0.1N HCl ^b | |
| F. Lobelia | | | | | F. Titration, 0.1N HCl ^b | |
| G. Nux vomica | | | | | G. Titration, 0.1N HCl ^{b,d} | |
| Brucine | Duolite CS-101 (NH ₄ ⁺) | | | 0.1M NH ₄ Cl | | (830) |
| Caffeine | Amberlite IRC-50 (H ⁺) | | | 6N HCl | | (829) |
| Cinchona bark* | Zeo-Karb ¹ (H ⁺) Duolite C-10 | 0.1N H ₂ SO ₄ | Dilute NaOH | Alcohol | | (24,25) (121) |
| Cinchona extract | Duolite C-10 (NH ₄ ⁺) | 46% Alcohol | (1) 46% alcohol, (2) water, (3) basic NH ₄ OH-NH ₄ Cl buffer | 5% NH ₄ OH in 95% alcohol | Titration, 0.1N HCl ^d | (120) |

| | | | | | | |
|----------------------------------|---|-------------------------------------|-------|---|---------------------------------------|----------------|
| Ephedrine | Amberlite IRC-50 (H ⁺) | | | 6N HCl | | (829) |
| Morphine sulfate | Amberlite IRC-50 Dowex 50 (H ⁺) | Alcohol Water | Water | 0.1N HCl Aqueous NaOH | Colorimetric, Radu- lescu reaction | (361) (768) |
| Morphine | Amberlite IRC-50 Amberlite IRC-50 (H ⁺) | Methanol | | 0.1N HCl 1N HCl or 4N NH ₄ OH in methanol | Spectrophotometric | (1) (321) |
| Morphine sulfate | Amberlite IR-120 | Water | | 4N NH ₄ OH in methanol | | (321) |
| Nicotine | Duolite CS-101 (NH ₄ ⁺) | | | 0.1M NH ₄ Cl | | (830) |
| Quinine | Amberlite IRC-50 Amberlite IRC-50 | Absolute alcohol | | 6N HCl Ammonia satu- rated alcohol | Polarimetric | (829) (648) |
| Quinine hydro- chloride | Duolite C-10 | 30% Alcohol | | 10% NH ₄ OH in 95% alcohol | | (120) |
| Quinine sulfate | Zeo-Karb [†] (H ⁺) | 0.1N H ₂ SO ₄ | Water | Alcoholic NaOH | | (512) |
| Strychnine | Duolite CS-101 (NH ₄ ⁺) | | | 0.1M NH ₄ Cl | | (830) |
| Sympathomime- tic amine salts | Amberlite IRC-50 | Water | Water | 75% Alcohol | Titration, 0.1N HCl ^d | (779) |

* In extracts, fluidextracts, tinctures, etc.

[†] Potentiometrically.

• To bromophenol blue indicator end point.

^d To methyl red indicator end point.

• Method of preparation of totaquine.

[†] Sulfonated coal.

Table 27

Summary of the application of ion exchangers in medicine with the example of Amberlite exchangers (CE = cation exchanger, AE = anion exchanger)

| <i>Stage of application</i> | <i>Field of application</i> | <i>Exchanger type</i> |
|--|--|---|
| Introduced applications | ulcers, hyperacidity | weak base AE; Amberlite IRP-58 |
| | adsorption of toxins, infantile diarrhea | weak base AE; Amberlite IRP-58 plus kaolin |
| | elimination of sodium, hypertension, and cardiac edema | weak acid CA; Amberlite IRP-64 and Amberlite IRP-88 |
| | elimination of potassium, kidney problems | weak acid CA; Amberlite IRP-64 |
| | tuberculosis; PAS-carrier | weak base AE; Amberlite IRP-58 as carrier for p-aminosalicylic acid |
| | diagnostic indicator of gastric acid | weak acid CE; Amberlite IRP-64 salt of Azure Blue |
| | dietary control, allergy, cough suppression | strong acid EC; Amberlite IR-120 as salt with alkaloids, antihistamine, amphetamine |
| | tablet disintegrator | K-form of weak acid CE; Amberlite IRP-88 |
| | vaginitis | weak acid CE; Amberlite IRP-64 |
| | vitamin B-12 stabilization | weak acid CE; Amberlite IRP-64 |
| | pruritis | strong base AE; Amberlite XE-235 |
| | ointment additive | mixture of all exchangers; Amberlite XE-87 |
| | Clinical investigations | against obesity |
| as laxative | | weak acid exchangers with little cross-linking, which swell in the intestine |
| as sustained-release hypertensive agents | | nitrate and nitrate forms of AE |

Blood therapy

decalcifying of blood
normalization of blood
and transfusion of pre-
served blood

Amberlite 200
mixture of all exchanger types

extracorporeal elimination
of K^+ and NH_4^+ in
hepatic coma

Amberlite 200

removal of toxins: for ex-
ample, elimination of
aspirin and barbiturates
in extracorporeal trans-
fusion

Amberlite IRA-900

artificial kidneys

Planned appli-
cations

elimination of common
salt

Amberlite IR-120

all-purpose antitoxin

types will low degree of cross-
linking

aspiring stabilization and
buffering

aspirin salt of Amberlite IRP-58

redox control

Amberlite XE-239

tablet drying

dry Amberlite types as drying
agents

surgical dressings, gauze

ion exchange fibers for pH con-
trol and as drug substitutes

Cuadro 2-1. Intercambiadores celulósicos de iones†

| Intercambiadores de aniones | Grupo ionizable | mEq/g* |
|------------------------------|---|---------|
| Celulosa AE | Aminoetilo $-O-CH_2-CH_2-NH_2$ | 0.3-1.0 |
| Celulosa DEAE | Dietilaminoetilo $-O-CH_2-CH_2-N(C_2H_5)_2$ | 0.1-1.1 |
| Celulosa TEAE | Trietilaminoetilo $-O-CH_2-CH_2-N(C_2H_5)_3$ | 0.5-1.0 |
| Celulosa GE | Guanidoetilo $-O-CH_2-CH_2-NH-C(=NH)-NH_2$ | 0.2-0.5 |
| Celulosa PAB | <i>p</i> -Aminobencilo $-O-CH_2-C_6H_4-NH_2$ | 0.2-0.5 |
| Celulosa ECTEOLA | Trietanolamina acoplada con celulosa mediante cadenas de glicerilo y poliglicerilo. Grupos mixtos | 0.1-0.5 |
| Celulosa BD | Celulosa DEAE benzoilada | 0.8 |
| Celulosa BND | Celulosa DEAE benzoilada y naftoilizada | 0.8 |
| Celulosa PEI | Polietilimina adsorbida a la celulosa o celulosa débil fosforilada | 0.1 |
| Intercambiadores de cationes | Grupo ionizable | mEq/g* |
| Celulosa CM | Carboximetilo $-O-CH_2-COOH$ | 0.5-1.0 |
| Celulosa P | Fosfato $-O-P(=O)(OH)_2$ | 0.7-7.4 |
| Célula SE | Sulfoetilo $-O-CH_2-CH_2-S(=O)_2-OH$ | 0.2-0.3 |

* La capacidad acidobásica está indicada por el fabricante.



Bacilo Gram (+)
Leuconostoc sp.

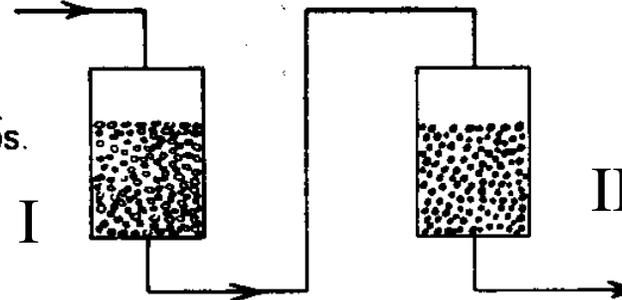
Lactosa: disacárido (gluc-
galact.)

↓
ATP, gluconatos-Ca ↓

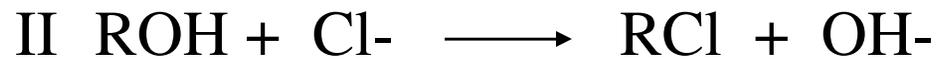
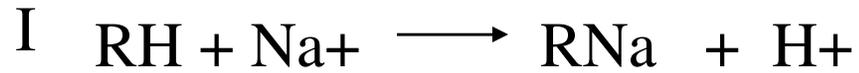
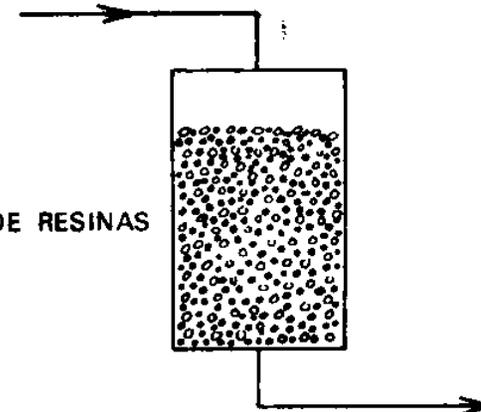
Leuconstoc mesenteroides

Azucarantos de Ca.

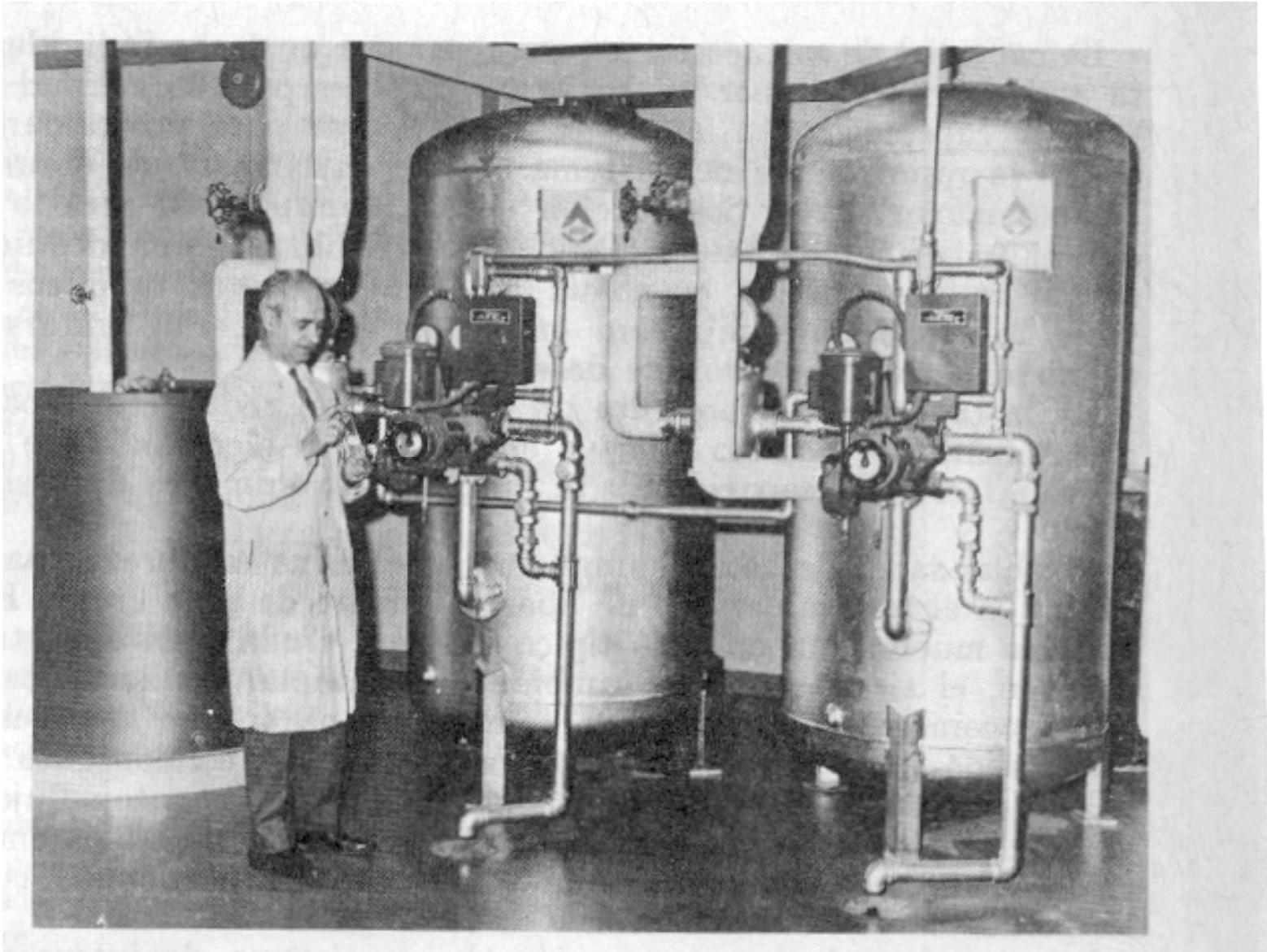
(a) INTERCAMBIADOR CONVENCIONAL DE CATION-ANION DE DOS LECHOS.



(b) DESMINERALIZACION POR ETAPAS MULTIPLES CON MEZCLA DE GRANOS DE RESINAS DE CATIONES Y DE ANIONES.



\longrightarrow H_2O
 \longrightarrow $microS/cm$
 < 1



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Tabla de variación de especies en intercambio iónico

| | | | | | | | |
|-------|----------------|-----|----------------|-----|-----------|-----|------------|
| | zRH | $+$ | $M(z+)$ | $=$ | RzM | $+$ | $zH+$ |
| Inc | mC_i | | $C_oV_o = n_o$ | | | | |
| Equil | $\approx mC_i$ | | $C_oV_o(1-f)$ | | fC_oV_o | | 10^{-pH} |

$$\text{Log } (f/1-f) = f \text{ (pH)}$$

$$K_I = \frac{(fC_oV_o/m) (10^{-zpH})}{(C_i)^z (C_o(1-f))}$$

$$\text{Log } (f/1-f) = \log K + z \log C_i + \log(m/V_o) + zpH$$

f = fracción intercambiada = (n_{int}/n_o)

C_i = capacidad científica de la resina mmolH/g

Condición operatoria $mC_i \gg C_oV_o$ pH impuesto.

Trazar las graficas $\log (f/1-f)$ para el sodio y el calcio y plantear Una posible separacion a pH controlado con resina cationica fuerte Amberlita IR-120 o Dowex 50 (ambas sulfónicas fuertes tipo RH):

Datos:

$$K_{\text{Na,H}} = 1.61$$

$$K_{\text{Ca,H}} = (2.28)^2$$

$$C_i = 5 \text{ mmol/g}; C_o = 0.01 \text{ M}; V_o = 25 \text{ mL}; m = 2.5 \text{ g.}$$

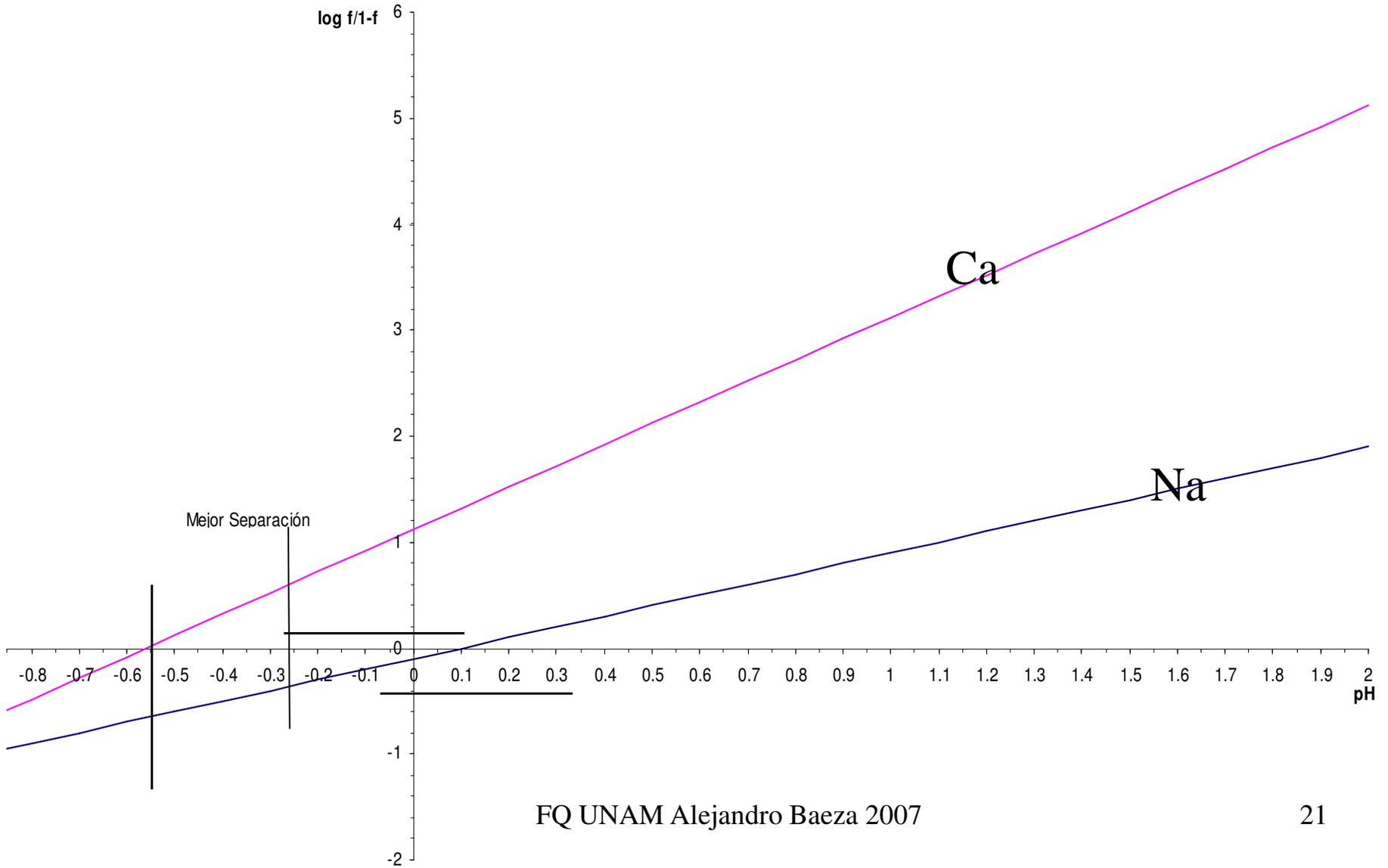
Tabla 6.1 A. Ringbom

“Formación de complejos en Química Analítica”

Editorial Alhambra. 1979. Pág. 234

$$\mathbf{Log (f/1-f) = log K + zlogCi + log(m/Vo) + zpH}$$

graficas log (f/1-f) para el sodio y el calcio



$C_i = 5 \text{ mmol/g}$; $C_o = 0.01 \text{ M}$; $V_o = 25 \text{ mL}$; $m = 2.5 \text{ g}$,

$$\text{Log } (f/1-f) = \log K + z \log C_i + \log(m/V_o) + z \text{pH}$$

Para Na

$$\log (f/1-f) = 0.2 + 0.7 \cdot -1 + \text{pH} = -0.1 + \text{pH}$$

Para Ca

$$\log (f/1-f) = 0.72 + 1.4 \cdot -1 + 2\text{pH} = 1.12 + 2\text{pH}$$

